Volume 50

2004

Didier Andrivon¹ Roselyne Corbière¹, Lionel Lebreton¹, Fabian Pilet¹, Josselin Montarry¹, Roland Pellé², Daniel Ellissèche²

¹ INRA, UMR INRA-ENSAR BiO3P, Domaine de la Motte, BP 35327, F-35653 Le Rheu Cedex, France; ² INRA, UMR INRA-ENSAR APBV, Keraiber, F-29260 Ploudaniel, France. *Author for correspondence: D. Andrivon, e-mail:* andrivon@rennes.inra.fr

HOST ADAPTATION IN *PHYTOPHTHORA INFESTANS*: A REVIEW FROM A POPULATION BIOLOGY PERSPECTIVE

ABSTRACT

Phytophthora infestans behaves in natural and agricultural ecosystems as a biotrophic pathogen, although it can be cultured on artificial media. Pathogenicity and host adaptation are therefore essential traits to understand its biology and to come up with durable, efficient management of late blight. The present review focuses on adaptation to host species and host cultivars, and to both qualitative and quantitative types of resistance. It also discusses some of the patterns and population mechanisms involved in this adaptation, such as selection, genetic drift and migration. This highlights the need for an in-depth analysis of each local situation to accurately describe and understand the mechanisms responsible for observed population displacements.

Key words: aggressiveness, host resistance, late blight, non-host resistance, specificity, virulence

INTRODUCTION

Phytophthora infestans, which causes late blight of Solanaceae, is often described as a highly variable pathogen. When applied to pathogenicity traits, this variability is regarded as the major cause for the breakdown of cultivar resistances used to control the disease. Consequently, over the last seventy years, a large corpus of research has dealt with the characterization of pathogenicity in *P. infestans* isolates or populations world-wide. Its primary aim has been to make a better use of the resistance sources available to potato breeders and growers, by choosing them according to the pathogenicity features prevalent in local *P. infestans* populations.

Different types of pathogenicity features must be considered in species which, like *P. infestans*, can interact with their hosts at various levels of specificity. Host specificity at the genus/species or at the cultivar

Communicated by Ewa Zimnoch-Guzowska

level allow to define the host range and physiological races of the pathogen, while the quantitative assessment of the disease induced in susceptible hosts is a major, but completely different component of pathogenicity (see for a discussion Andrivon 1993). We therefore investigated the extent of pathogenic variation present in "old" and "new" populations of *P. infestans* (*sensu* Spielman *et al.* 1991) at the different specificity levels, through a review of some of the published data.

The introduction, beginning in the mid-to late 1970s, of a number of pathogen genotypes into Europe, and subsequently to most potato cropping areas in the world (Spielman et al. 1991, Fry et al. 1992, 1993, Goodwin et al. 1994), led to a rapid displacement of local isolates (called "old populations" by Spielman et al. 1991) by newly introduced ones. The extreme speed of this population shifts, and their world-wide occurrence despite the variation in the characteristics of immigrant genotypes, led to further concern about the extent and sources of variability present in *P. infestans*. Three main factors can be advocated to account for the evolutionary success of the immigrant strains over previously established populations: increased pathogenicity, in terms of host range and of aggressiveness to each major host (Fry et al. 1992, Day and Shattock 1997, Kato et al. 1997, Miller et al. 1998); increased fitness, through higher adaptability (Fry et al. 1992), a wider range of ecological competence (e.g. Mizubuti and Fry 1998), or better survival as a consequence of the possible occurrence of the sexual stage (Fry *et al.* 1989, 1992, Andrivon 1995); and higher gene flow, either because of migrations or of sexual reproduction (Goodwin et al. 1998).

Although much work has been dedicated to the identification of population structures of *P. infestans* in North America (e.g. Fry *et al.* 1992, Goodwin et al. 1992, 1994, 1998), Europe (e.g. Fry et al. 1991, Andrivon et al. 1994, Drenth et al. 1994, Day and Shattock 1997, Lebreton et al. 1998, Carlisle et al. 2001, Cooke et al. 2003) and other parts of the world (e.g. Goodwin et al. 1994, Koh et al. 1994, Forbes et al. 1997, Reis et al. 2003), we are still a long way from a firm assessment of the implications of the three above-mentioned mechanisms in the current set-up and recent evolution of populations of the fungus in the different geographical and ecological situations where the pathogen prospers. One of the major reasons for this is the fact that most recent population research has focused on selectively neutral markers, but much less on pathogenicity features considered from a population biology perspective. Furthermore, these studies have shown that the characteristics of the groups of genotypes of the pathogen present in the various parts of the world, while different from those of the original set of clones (US-1 and related sub-clones; Goodwin et al. 1994, 1998), are also markedly distinct from one another. Populations present in North America are usually quite simple, with one genotype dominating at each location in a given year (Goodwin et al. 1998), whereas the setup of European populations is often much more complex, with a number of genotypes coexisting at any single time (Drenth et al. 1993, Lebreton and Andrivon 1998, Lebreton et al. 1998, Cooke et al. 2003). Finally, while late blight resurgence is a fairly recent phenomenon in North America (Fry and Goodwin 1997), the disease has been prevalent in Europe and in several other parts of the world for the past 150 years, with years without severe outbreaks the exception more than the rule (e.g. Large 1953, 1956, Schöber 1987). This much greater prevalence of the disease in Europe is directly translated in terms of pesticides used to control late blight. While growers in the Columbia basin of Washington and Oregon would spray an average of 2.5 times against *P. infestans* in a year of low incidence of the disease (such as 1994) and up to 10 times in a severe blight year, such as 1995 (Johnson et al. 1997), potato crops in western Europe would typically be sprayed an average of 6-8 times against blight, and up to 15–20 times in severe blight years and susceptible cultivars (Schepers 2003). All together, the large discrepancies in population composition and epidemic potential make generalizations from one population to others problematic.

The aims of this paper are to review the information currently available about patterns of adaptation to hosts in populations of *P. infestans*, and about the evolutionary mechanisms related to these patterns. The goal is to provide keys to explain – and, if possible, predict – future changes in pathogenicity features at the population level, which are needed for a better and more durable management of genetic resources for resistance to late blight. Because most of the data available concern either North American or European populations, the review is focused on these two groups of populations.

ADAPTATION TO HOST GENUS/SPECIES

P. infestans is known to be pathogenic to at least forty species of Solanaceae (Turkensteen 1978). Host specificity is of pathological, but also of evolutionary significance, because the possibility for infecting more than one host determines to a large extent the availability of "green bridges" during the pathogen's life cycle. These are critical in maximizing survival opportunities in species with very low saprophytic abilities, such as *P. infestans* (see Andrivon 1995 for a review), and probably condition the extent of gene flow between isolates. Host specificity may also have led to a speciation event between *P. infestans* and *P. mirabilis*, two species giving rise to fertile hybrids (Goodwin and Fry 1994), morphologically indistinguishable from one another (Galindo and Hohl 1985), but with mutually exclusive host ranges. This separation of host ranges explains the reproductive isolation of *P. infestans* and *P. mirabilis* in nature, which in turn probably led to the accumulation of genetic differences detectable in current collections (Goodwin, personal communication). P. mirabilis was thus considered either as a variety (Servin 1958) or a forma specialis of P. infestans (Möller et al. 1993), or regarded as a valid species (Galindo and Hohl 1985, Goodwin and Fry 1994, Goodwin and others, personal communication). Similar speciation patterns have been described in recent years in South America (Adler *et al.* 2002). They involve sympatric wild and/or cultivated hosts, which points to a selective advantage to host specialisation in habitats where a number of potential hosts are present (Lapchin 2002). However, this general trend towards specialisation (i.e. restriction of host range) is sometimes reverted, as shown by the recent discovery in the Netherlands of isolates overcoming the resistance of *Solanum nigrum*, until then regarded as a non-host for *P. infestans* (Flier *et al.* 2003a).

The specialisation process does not always lead to species individualisation. Many authors observed isolates more specifically adapted to either potato (*Solanum tuberosum*) or tomato (*Lycopersicum esculentum*), both in "old" (e.g. Berg 1926, Small 1938) and in "new" populations (Legard *et al.* 1995, Lebreton *et al.* 1998, 1999, Oyarzun *et al.* 1998, Reis *et al.* 2003), but specificity was never restrictive enough to warrant the "forma specialis" denomination. The initial adaptation of any isolate can be reverted by repeated passages through the other host. Although this process is unlikely to be of significance in agricultural practice, since isolates initially adapted to one host would be out-competed on the other host, it shows a high level of genetic plasticity in *P. infestans* regarding pathogenicity determinants.

Two isolates of *P. infestans* collected from tomato in southwestern Europe and characterized by isozyme alleles and mitochondrial haplotypes as "old" populations showed different genetic fingerprints (Lebreton and Andrivon 1998), which suggests that the higher genetic diversity of the pathogen observed today on tomato in this area might have existed before the introduction of "new" genotypes. In many "new" populations, differences in the frequency distribution of genetic features other than pathogenicity (such as mating types, mitochondrial DNA haplotypes, isozyme alleles, or nuclear DNA fingerprints) between collections of isolates made on potato and on tomato is the rule rather than the exception (Lebreton and Andrivon 1998, Oyarzun et al. 1998). Interestingly, isolates present on tomato generally belong to simpler races than those collected on potato (Deahl et al. 1993, Lebreton and Andrivon 1998). However, there is no consistent association on a world-wide basis between genetic markers and adaptation to one or the other host. For instance, A2 isolates are more frequently found on tomato than on potato in France (Lebreton and Andrivon 1998), but are restricted to potato in several South American countries, such as Brazil (Brommonschenkel 1988, Reis et al. 2003).

ADAPTATION TO HOST CULTIVARS

Virulence to *R*-genes

The existence of race-specific resistance genes in Solanum tuberosum and of matching physiological races in *P. infestans* have been recognised since the 1940s and extensively investigated since the early 1950s (see Wastie 1991 for a review). The initial 11 *R*-genes described originate from the Mexican species Solanum demissum, but similar genes exist in many other tuber-bearing Solanum species (Hawkes 1958, Rivera-Peña 1990, Tooley 1990). Some of these species, including S. *bulbocastanum*, have been considered as promising sources of highly efficient R-genes, and the corresponding R-genes have been mapped and/or cloned (Song et al. 2003, Van der Vossen et al. 2003). However, both the RB / Rpi-blb1 genes recently cloned from S. bulbocastanum and *R1* from *S. demissum* (Ballvora *et al.* 2002) are genetically similar to many known – and defeated – R–genes which belong to the NBS–LRR (nucleotide binding site-leucine rich repeat) class. This suggests that all can be defeated rapidly by new races of *P. infestans* if deployed in commercial cultivars.

Because deployment strategies of R-genes are an obvious factor shaping the changes in race frequencies, complex races were selected in "old" populations wherever popular potato cultivars carried combinations of R-genes, such as in Great Britain (Malcolmson 1969, Shattock *et al.* 1977). In some instances, the introduction of "new" populations led to a marked increase in the complexity of races (Deahl *et al.* 1993, Drenth *et al.* 1994). However, a comparative analysis of race structure characteristics in "old" and "new" populations of P. *infestans* collected world-wide and surveyed with the same set of differential clones showed no consistent trend towards an increase in virulence complexity or virulence diversity in the most recent populations (Andrivon 1994a).

Part of the discrepancy between regions might be due to the fact that most of the virulences present in the isolates now recovered outside Mexico do not reflect local selection, but selection in Mexico prior to migration. A number of virulences present nowadays in Europe (such as virulences to R7, which is very common, and virulences to R5, R6 and R8, which are less frequent) match resistance genes which have never been used in European commercial cultivars. These virulences can be regarded as "fossil" traits, once selected for in central Mexico where the matching R-genes exist in nature, and maintained in isolates after their migration, as no genetic mechanism existed to remove them. Accumulation of "fossil virulences" has been postulated in rust pathogens of cereals, although in this case the original selection was exerted locally (Andrivon and de Vallavieille–Pope 1995). From an evolutionary per– spective, this implies the absence of a fitness penalty associated with unnecessary virulences, making most virulences behave as selectively neutral markers when resistance genes are not used, which is the case

19

now in several parts of Europe (Andrivon *et al.* 1994) and in North America (Fry and Goodwin 1997). Indeed, there seems to be no correlation between fitness and race complexity in "new" European populations (Schöber and Turkensteen 1992, Pilet 2003). It is therefore difficult to attribute the success of immigrant genotypes in displacing former clones to their higher virulence complexity, because (i) not all migrant genotypes belonged to races more complex than the clones they displaced, and (ii) no immediately identifiable fitness benefit can be associated with this increase, when it occurred.

Aggressiveness

P. infestans isolates have been shown to vary largely in their aggressiveness towards potato cultivars. This variation is not related to physiological races (e.g. Jeffrey et al. 1962, Denward 1967, Caten 1974), and can be detected both in controlled conditions and in the field (e.g. Tooley and Fry 1985, Tooley et al. 1986, Day and Shattock 1997). Aggressiveness can decrease during repeated subculturing on artificial media, but it can be restored *via* inoculation of living plant material (Jeffrey *et al.* 1962, Jinks and Grindle 1963). In several experiments, aggressiveness remained stable over successive transfers to potato plants (Caten 1974). Furthermore, specific components of aggressiveness have been detected repeatedly (e.g. de Bruyn 1947, Jeffrey et al. 1962, Jinks and Grindle 1963, Caten 1974, Carlisle et al. 2002, Corbière et al. 2002), reflecting the fact that each isolate usually grows better on the variety it was recovered from than on other varieties with the same R-genes. However, recent work with populations sampled at the same locations on cultivars free of R-genes, but expressing different levels of partial resistance, consistently revealed higher average agressiveness towards both susceptible and partially resistant cultivars in populations sampled from susceptible rather than from partially resistant hosts (Pilet 2003).

Evidence is accumulating to show a higher aggressiveness in isolates belonging to "new" populations than in their "older" counterparts (Day and Shattock 1997, Kato *et al.* 1997, Flier and Turkensteen 1999). How– ever, because of methodological limitations, these data should be inter– preted with caution. Aggressiveness among isolates is a composite of many traits, and is thus difficult to measure accurately. Variation is characteristic of most of aggressiveness components (e.g. latent period, infection efficiency, sporulation), but is not always directly correlated with disease progress in the field (Spielman *et al.* 1992). Therefore, comparisons made on single components might not accurately describe actual differences in global aggressiveness between isolates. Further– more, "old" and "new" isolates being compared may not have been sub– jected to the same number of transfers on artificial media. Finally, the extensive variation present among "new" isolates of *P. infestans* some– times overlaps the range of differences between "old" and "new" isolates (Schepers 1998). Indeed, US–1 remains a very destructive pathogen in some tropical highland areas, such as Ecuador (Oyarzun *et al.* 1998), or in temperate conditions in South Africa (McLeod *et al.* 1998).

While blight has undoubtedly gained importance in North America over the last decade (Fry and Goodwin 1997), no consistent trend towards a worsening of the disease appears from long-term surveys of blight incidence and severity in Europe. This discrepancy might be explained by the fact that, contrary to what happens in most parts of North America, blight is observed every year in western Europe, where the pathogen usually finds very favourable climatic conditions (mild, humid climate). However, it is noteworthy that not all years since the introduction of "new" strains have been severe blight years in Europe. For instance, Hardwick and Turner (1996) reported that blight incidence and severity were low in England and Wales in the early the 1990s, although only "new" genotypes were present (Day and Shattock 1997). A similar situation occurred in France in 1989 and 1990 (Duvauchelle 1993). The use of fungicides for blight control in Europe was 15-40% higher in 1997 (severe blight) than in 1996 (moderate blight) (Schepers 1998), although there was no evidence of major changes in the pathogen population between the two years. Overall, the incidence and severity of blight seems to be much better correlated (at least in Europe) with the climatic conditions prevalent in a given year than with the type of populations present. Indeed, disease forecasting systems, despite being based on experimental data relating to "old" isolates, still work pretty accurately, the major factor conditioning their performance being the quality of input meteorological data (Hansen 1998).

PATHOGENICITY, ADAPTATION, AND FITNESS

Genetic diversity and adaptability

"New" populations of the pathogen are constituted of a larger number of genotypes, and are usually genetically more diverse (at least for neutral markers) than were "old" populations (Goodwin et al. 1994). However, it is not known how this higher diversity is translated in terms of adaptability. As discussed above, there is no unequivocal evidence that new populations are always more pathogenic than their former counterparts. While there is some evidence for adaptation to a broader range of climates in "new" lineages (Mizubuti and Fry 1998), these data are not unequivocal. For instance, over a range of temperatures, US-1 sporulated and germinated on average more abundantly than US-7 and US-8 in controlled experiments (Mizubuti and Fry 1998), although these authors concluded that new genotypes were fitter than members of the US-1 lineage from measurements of incubation period and/or lesion size. Flier et al. (2003b) also showed that US-1 was more pathogenic than US-8 on some European potato cultivars, and that "new" European isolates exhibited a wide range of variability concerning the pathogenicity to these cultivars. Finally, recent work done in France and Ecuador (Pilet 2003) has revealed the existence of extensive variability for aggressiveness in isolates belonging to the same AFLP (amplified fragment length polymorphism) pattern.

Survival

The formation of long lasting oospores is an obvious advantage for survival in an organism with saprophytic capacities as low as those of P. infestans (Fry et al. 1989, Andrivon 1995). This advantage is even greater in parts of the world with extreme climates (either hot summers, such as in Mexico, or very cold winters, such as in Canada, Poland or Scandinavia). Not surprisingly, the most convincing evidence for involvement of sexual spores in the epidemiology of late blight comes just from such areas (Sujkowski et al. 1994, Goodwin et al. 1995, Andersson et al. 1998). In areas with less contrasted seasons, the opportunities for asexual survival of the pathogen in infected tubers (volunteers, refuse piles) are probably high enough to ensure that the largest part of the primary inoculum is of asexual origin, even though oospores might be present. The difficulty is then to detect the fraction of the inoculum derived from oospores in the global populations of isolates. Indeed, refuse piles remain the major primary infection sources for commercial potato crops in Flevoland, the Netherlands, despite the occurrence in the immediate vicinity of allotment gardens where both mating types and oospores are present (Zwankhuizen et al. 1998).

A major consequence of the poor survival of *P. infestans* outside its host and of the limited number of situations where oospores play a significant epidemiological part to date is the structural lability noticed in many local populations of the fungus over a series of years (Drenth et al. 1993, Andrivon 1994a, Goodwin et al. 1998, Lebreton et al. 1998, Zwankhuizen et al. 1998). This makes impossible to predict the population structure in a given year from the knowledge of its structure in former years (Fry and Goodwin 1997, Goodwin et al. 1998, Lebreton et al. 1998), at the same time indicating a major role of founder effects in shaping population structures over time (Fry et al. 1992, Andrivon 1994a, Fry and Goodwin 1997, Lebreton et al. 1998). These observations strongly suggest that pathogenic fitness is one, but probably not the main, of the components of global fitness in *P. infestans* genotypes, and that chance and survival ability are more critical than pathogenicity in long term evolutionary success. In this respect, the observation that in Brittany, late blight epidemics are often started by very simple races, usually found mainly on tomatoes and present late in the season but less pathogenic to potato, and that the major, complex potato races only develop later (Andrivon 1994b), is significant, because it illustrates the fact that the highest pathogenicity is not a prerequisite for persistence over time, and might even hinder it. This resembles wild host-parasite systems, where balanced pathogenicity commonly develops (Bull 1994, Lapchin 2002).

SOME CONCLUSIONS

This review confirms that extensive variation exists in both "old" and "new" populations of P. infestans for all components of pathogenicity, but also outlines the fact that pathogenicity is only one element of the story when attempting to understand population structures in this pathogen. Although the evidence exists that "new" populations might be on average more virulent (i.e. include more complex races) and more aggressive than their former counterparts, this trend does not refer equally to all situations, and exceptions can be easily found in the available data. Furthermore, the large influence of conditions prevalent during epidemic development, but even more during survival, result in strongly pronounced drift effects, which means that the actual fitness of a genotype (i.e. its contribution to the next generation or to the population in the following year), is only partially dependent on the pathogenicity of this genotype. This has two major consequences for explaining and predicting changes in population structures of the late blight pathogen: first, the extent to which the changes of pathogenicity affect population structures cannot be evaluated on a general and uniform basis, but needs to be assessed for a particular situation; second, it is very difficult to predict population changes based only on pathogenicity features. The metapopulation structure of *P. infestans* populations on local and regional scales (Andrivon et al. 1994, Lebreton and Andrivon 1998) make this prediction even more difficult, because of the random nature of founding events and of the poor quantitative assessment of migration rates between patches. Until comprehensive models, including random (or more likely frequency-dependent) extinction events during survival stages, possibilities for switching hosts, and pathogenic fitness on each host, are developed, predicting population structures of *P. infestans* will remain largely a guess. Further work is therefore needed to better appreciate and quantify the amount of gene flow, the modalities of extinction and the impact of long-lasting oospores on survival and population structures in the long term, which are the absolute prerequisites for the development of such comprehensive models.

REFERENCES

- Adler N., Chacon G., Forbes G., Flier W. 2002. Phytophthora infestans sensu lato in South
- America population G., FORDES G., FHER W. 2002. Phytophthora infestans sensu lato in South America population substructuring through host specificity. (In:) Late blight manag-ing the global threat. Lizarraga C. (ed.). GILB-CIP, Lima, Peru: 13-17.
 Andersson B., Sandström M., Strömberg A. 1998. Sexual reproduction of Phytophthora infestans on potato in Sweden. (In:) Proc. 2nd workshop European network for develop-ment of an integrated control strategy of potato late blight, PAV Special Report 3. Schepers H. Bouma E. (eds): 92-96 Schepers H., Bouma E. (eds): 92-96.
- Andrivon D. 1993. Nomenclature for pathogenicity and virulence: the need for precision. Phytopathology 83: 889-890.

Andrivon D. 1994a. Race structure and dynamics in populations of *Phytophthora infestans*. Can. J. Bot. 72: 1681–1687.

Andrivon D. 1994b. Races of Phytophthora infestans in France, 1991-1993. Potato Res. 37: 279 - 286.

Andrivon D. 1995. Biology, ecology and epidemiology of the potato late blight pathogen *Phy-tophthora infestans* in soil. Phytopathology 85: 1053–1056.
 Andrivon D., Béasse C., Laurent C. 1994. Characterization of isolates of *Phytophthora infestans* collected in northwestern France from 1998 to 1992. Plant Pathol. 43: 471–478.

Andrivon D., Vallavieille-Pope C. de. 1995. Race diversity and complexity in selected popula-

Andrivon D., Vanaviene-rope C. de. 1955. Race diversity and complexity in selected populations of fungal biotrophic pathogens of cereals. Phytopathology 85: 897–905.
 Ballvora A., Ercolano M.R., Weiss J., Meksem K., Bormann C.A., Oberhagemann P., Salamini F., Gebhardt C. 2002. The R1 gene for potato resistance to late blight (*Phytoph-thora infestans*) belongs to the leucine zipper/NBS/LRR class of plant resistance genes.

- Plant J. 30: 361–371.
- Berg A. 1926. Tomato late blight and its relation to late blight of potato. W. Va. Agric. Exp. Stn Bull. 205
- Brommonschenkel S.H. 1988. Pathogenicity, compatibility, cytogenetics and isoenzyme pat-terns of Brazilian isolates of *Phytophthora infestans* (Mont.) de Bary. M.Sc. thesis. Universidade Federal de Viçosa, Viçosa, Brazil.

Bull J.J. 1994. Perspective: virulence. Evolution 48: 1423–1437.
Carlisle D., Cooke L.R., Brown A.E. 2001. Phenotypic and genotypic characterisation of Northern Ireland isolates of *Phytophthora infestans*. Eur. J. Plant Pathol. 107: 291–303.
Carlisle D., Cooke L.R., Watson S., Brown A.E. 2002. Foliar aggressiveness of Northern Ire-land isolates of *Phytophthora infestans* on detached leaflets of three potato cultivars. Distribution of the potato cultivars. Plant Pathol. 51: 424-434.

Caten C.E. 1974. Intra-racial variation in *Phytophthora infestans* and adaptation to field resistance for potato blight. Ann. Appl. Biol. 77: 259–270.
Cooke D.E.L., Young V., Birch P.R.J., Toth R., Gourlay F., Day J.P., Carnegie S.F., Duncan

- J.M. 2003. Phenotypic and genotypic diversity of *Phythophthora infestans* populations in Scotland (1995–1997). Plant Pathol. 52: 181–192.
- Corbière R., Trémoulu B., Lucas J.M., Pellé R., Ellissèche D., Andrivon D. 2002. Components of partial resistance to late blight in ancient potato cultivars. (In:) 15th Trien. Conf. EAPR. Hamburg, Germany, 14–19.07.02: 102.
 Day J.P., Shattock R.C. 1997. Aggressiveness and other factors relating to displacement of
- populations of *Phytophthora infestans* in England and Wales. Eur. J. Plant Pathol. 103: 379–391.
- Deahl K.L., Inglis D.A., Muth S.P. de 1993. Testing for resistance to metalaxyl in Phytophthora infestans from northwestern Washington. Am Potato J. 70: 779–795. De Bruyn H.G.L. 1947. Het rassenproblem bij *Phytophthora infestans*. Vakbl. Biol. 27:
- 147-152.

Denward T. 1967. Differentiation in *Phytophthora infestans*. I. A comparative study of eight different biotypes. Hereditas 58: 191–220.

- Drenth A., Goodwin S.B., Fry W.E., Davidse L.C. 1993. Genotypic diversity of *Phytophthora* infestans in the Netherlands revealed by DNA polymorphisms. Phytopathology 83: 1087-1092.
- Drenth A., Tas I.C.Q., Govers F. 1994. DNA fingerprinting uncovers a new sexually reproducing population of *Phytophthora infestans* in the Netherlands. Eur. J. Plant Pathol. 100: 97–107.

Duvauchelle S. 1993. Les caprices du mildiou. Phytoma - Def. Veg. 449: 46-47.

- Flier W.G., Bosch G.B.M. van den, Turkensteen L.J. 2003a. Epidemiological importance of Solanum sisymbriifolium, S. nigrum and S. dulcamara as alternative hosts for Phytoph-thora infestans. Plant Pathol. 52: 595–603.
- Flier W.G., Bosch G. B. M. van den, Turkensteen L. J. 2003b. Stability of partial resistance in potato cultivars exposed to aggressive strains of *Phytophthora infestans*. Plant Pathol. 52: 326-337.
- Flier W.G., Turkensteen L.J. 1999. Foliar aggressiveness of Phytophthora infestans in three potato-growing regions in the Netherlands. Eur. J. Plant Pathol. 105: 381-388. Forbes G.A., Escobar X.C., Ayala C.C., Revelo J., Ordoñez M.E., Fry B.A., Doucett K., Fry
- W.E. 1997. Population genetic structure of Phytophthora infestans in Ecuador. Phytopathology 87: 375-380.
- Fry W.E., Drenth A., Spielman L.J., Mantel B.C., Davidse L.C., Goodwin S.B. 1991. Population genetic structure of *Phytophthora infestans* in the Netherlands. Phytopathology 81: 1330-1336.
- Fry W.E., Goodwin S.B. 1997. Re-emergence of potato and tomato late blight in the United States. Plant Dis. 81: 1349-1357.

- Fry W.E., Goodwin S.B., Dyer A.T., Matuszak J.M., Drenth A., Tooley P.W., Sujkowski L.S., Koh Y.J., Cohen B.A., Spielman L.J., Deahl K.L., Inglis D.A., Sandlan K.P. 1993. Histor-ical and recent migrations of *Phytophthora infestans*: chronology, pathways, and impli-cations. Plant Dis. 77: 653-661.
 Fry W.E., Goodwin S.B., Matuszak J.M., Spielman L.J., Milgroom M.G., Drenth A. 1992. Population genetics and intercontinental migrations of *Phytophthora infestans*. Annu. Durbusched. 200, 1027–100.
- Population genetics and intercontinental inigrations of *Phytophenora infectance*. Final, Rev. Phytopathol. 30: 107–129.
 Fry W.E., Tooley P.W., Spielman L.J. 1989. The importance of the perfect stage of *Phytoph-thora infestans* from the standpoint of epidemiology and adaptation. (In:) Fungal diseases of the potato. CIP, Lima, Peru: 17–30.
 Galindo A.-J., Hohl H.R. 1985. *Phytophthora mirabilis*, a new species of *Phytophthora*. Culture 20: 97–96.
- Sydowia 38: 87-96.
- Goodwin S.B., Cohen B.A., Fry W.E. 1994. Panglobal distribution of a single clonal lineage of the Irish potato famine fungus. Proc. Nat. Acad. Sci. USA 91: 11591–11595. Goodwin S.B., Fry W.E. 1994. Genetic analyses of interspecific hybrids between *Phytoph*–
- Goodwin S.B., Fry W.E. 1994. Genetic analyses of interspectic hybrids between Thytoph-thora infestans and Phytophthora mirabilis. Exp. Mycol. 18: 20-32.
 Goodwin S.B., Smart C.D., Sandrock R.W., Deahl K.L., Punja Z.K., Fry W.E. 1998. Genetic change within populations of Phytophthora infestans in the United States and Canada during 1994 to 1996: role of migration and recombination. Phytopathology 88: 939–949.
- Goodwin S.B., Spielman L.J., Matuszak J.M., Bergeron S.N., Fry W.E. 1992. Clonal diversity and genetic differentiation of *Phytophthora infestans* populations in northern and cen-
- tral Mexico. Phytopathology 82: 955-961.
 Goodwin S.B., Sujkowski L.S., Dyer A.T., Fry B.A., Fry W.E. 1995. Direct detection of gene flow and probable sexual reproduction of *Phytophthora infestans* in northern North America. Phytopathology 85: 472–470. America. Phytopathology 85: 473–479.
- Hansen J.G. 1998. Availability and use of meteorological data for disease forecasting in Den-mark. (In:) Proc. 2nd workshop European network for development of an integrated con-trol strategy of potato late blight, PAV Special Report 3. Schepers H., Bouma E. (eds): 125 - 136.
- Hardwick N.V., Turner J.A. 1996. A survey of potato diseases in England and Wales 1993–1996. (In:) Abstracts, 13th Trien. Conf. EAPR. Veldhoven, Netherlands, 14–19 July 1996: 631-632.
- Hawkes J.G. 1958. Significance of wild species and primitive forms for potato breeding. Euphytica 7: 257-270.

Jeffrey S.I.B., Jinks J.L., Grindle M. 1962. Intraracial variation in Phytophthora infestans and field resistance to potato blight. Genetica 32: 323–338. Jinks J.L., Grindle M. 1963. Changes induced by training in *Phytophthora infestans*. Hered–

ity 18: 245-264.

Johnson D.A., Cummings T.F., Hamm P.B., Rowe R.C., Miller J.S., Thornton R.E., Pelter G.Q., Sorensen E.J. 1997. Potato late blight in the Columbia Basin: an economic analysis of the 1995 epidemic. Plant Dis. 81: 103–106.
 Kato M., Mizubuti E.S., Goodwin S.B., Fry W.E. 1997. Sensitivity to protectant fungicides

- and pathogenic fitness of clonal lineages of Phytophthora infestans in the United States.
- and pathogenic niness of cional inleages of *Layophenesic englished* a supervise a supervise
- Phytopathology 84: 922–927.
 Lapchin L. 2002. Host-parasitoid association and diffuse coevolution: When to be a generalist? Am. Nat. 160: 245–254.
 Large E.C. 1953. Potato blight forecasting investigation in England and Wales, 1950–52.

- Plant Pathol. 2: 1–15. Large E.C. 1956. Potato blight forecasting and survey work in England and Wales, 1953–55. Plant Pathol. 5: 39–52
- Lebreton L., Andrivon D. 1998. French isolates of Phytophthora infestans from potato and tomato differ in phenotype and genotype. Eur. J. Plant Pathol. 104: 583–594. Lebreton L., Laurent C., Andrivon D. 1998. Evolution of *Phytophthora infestans* populations
- in the two most important potato production areas of France during 1992-96. Plant Pathol. 47: 427-439
- Lebreton L., Lucas J.M., Andrivon D. 1999. Aggressiveness and competitive fitness of Phytophthora infestans isolates collected from potato and tomato in France. Phytopathology 89:679-686.

Legard D.E., Lee T.Y., Fry W.E. 1995. Pathogenic specialization in Phytophthora infestans: aggressiveness on tomato. Phytopathology 85: 1356-1361.

Malcolmson J.F. 1969. Races of *Phytophthora infestans* occurring in Great Britain. Trans. Brit. Mycol. Soc. 53: 417-423.

- McLeod A., Denman S., Denner F.D.N., Sadie A. 1998. Characterization of *Phytophthora* infestans populations in South Africa. (In:) 7th Intl. Cong. Plant Pathology Offered Pa-
- pers, Vol. 2: Abstract 2.2.57.
 Miller J.S., Johnson D.A., Hamm P.B. 1998. Agressiveness of isolates of *Phytophthora* infestans from the Columbia Basin of Washington and Oregon. Phytopathology 88: 190 - 197
- Mizubuti E.S.G., Fry W.E. 1998. Temperature effects on developmental stages of isolates from three clonal line ages of *Phytophthora infestars*. Phytopathology 88: 837–843. Möller E.M., de Cock A.W.A.M., Prell H.H. 1993. Mitochondrial and nuclear DNA restriction
- enzyme analysis of the closely related *Phytophthora* species *P. infestans*, *P. mirabilis* and *P. phaseoli*. J. Phytopathol. 139: 309–321.
- Oyarzun P.J., Pozo A., Ordoñez M.E., Doucett K., Forbes G.A. 1998. Host specificity of *Phy-tophthora infestans* on potato and tomato in Ecuador. Phytopathology 88: 265–271.
- Pilet F. 2003. Epidémiologie et biologie adaptative des populations de Phytophthora infestans dans des cultures pures et hétérogènes de variétés de pomme de terre. Ph.D. thesis. Ecole
- Nationale Supérieure Agronomique de Rennes, France, 157 p. Reis A., Smart C.D., Fry W.E., Maffia L.A., Mizubuti E.S.G. 2003. Characterization of isolates of *Phytophthora infestans* from southern and southeastern Brazil from 1998 to 2000. Plant Dis. 87: 896-900.
- Rivera-Peña A. 1990. Wild tuber-bearing species of Solanum and incidence of Phytophthora infestans (Mont.) de Bary on the western slopes of the volcano Nevado de Toluca. 5. Type of resistance to Phytophthora infestans. Potato Res. 33: 479-486.
- Schepers H.T.A.M. 1998. Epidemiological parameters in decision support systems for *Phy-tophthora infestans*. (In:) Proc. 2nd workshop European network for development of an integrated control strategy of potato late blight, PAV Special Report 3. Schepers H., Bouma
- E. (eds): 30-36.
 Schepers H.T.A.M. 2003. The development and control of *Phytophthora infestans* in Europe in 2002. (In:) Proc. 7th workshop European network for development of an integrated control strategy of potato late blight, PAV Special Report 9. Westerdijk C.E., Schepers H.T.A.M. (eds): 9–22.
- Schöber B. 1987. Phytophthora infestans (Mont.) de Bary: eine ständige Herausforderung seit 140 Jahren. Ber. Deutsch. Bot. Gesellsch. 100: 291-303.
- Schöber B., Turkensteen L.J. 1992. Recent and future developments in potato fungal pathology. Neth. J. Plant Pathol. 98, Suppl. 2: 73-83.
- Servin L. 1958. Especie de *Phytophthora* atacando a *Mirabilis jalapa*. Mem. Primer Congr. Nac. Entomol. Fitopatol., Mexico: 491–500.
 Shattock R.C., Janssen B.D., Whitbread R., Shaw D.S. 1977. An interpretation of the frequencies of host specific phenotypes of *Phytophthora infestans* in North Wales. Ann. Appl. Biol. 86: 249–260.
 Small T. 1938. The relation between potato blight and tomato blight. Ann. Appl. Biol. 25: 2021. 2020.
- 271-276.
- Song J.Q., Bradeen J.M., Naess S.K., Raasch J.A., Wielgus S.M., Haberlach G.T., Liu J., Kuang H.H., Austin-Phillips S., Buell C.R., Helgeson J.P., Jiang J.M. 2003. Gene RB cloned from *Solanum bulbocastanum* confers broad spectrum resistance to potato late blight. Proc. Natl. Acad. Sci. USA 100: 9128-9133.
 Spielman L.J., Drenth A., Davidse L.C., Sujkowski L.J., Gu W., Tooley P.W., Fry W.E. 1991.
- A second world-wide migration and population displacement of *Phytophthora infestans*? Plant Pathol. 40: 422-430.
- Spielman L.J., McMaster B.J., Fry W.E. 1992. Relationships between measurements of fitness and disease severity in *Phytophthora infestans*. Plant Pathol. 41, 317–324. Sujkowski L.S., Goodwin S.B., Dyer A.T., Fry W.E. 1994. Increased genotypic diversity via
- migration and possible occurrence of sexual reproduction of *Phytophthora infestans* in
- Poland. Phytopathology 84: 201–207. Tooley P.W. 1990. Variation in resistance to *Phytophthora infestans* among 21 Solanum verrucosum plant introductions. Am. Potato J. 67: 491–498. Tooley P.W., Fry W.E. 1985. Field assessment of fitness of isolates of *Phytophthora infestans*.
- Phytopathology 75: 982–988. Tooley P.W., Sweigard J.A., Fry W.E. 1986. Fitness and virulence of *Phytophthora infestans*
- isolates from sexual and asexual populations. Phytopathology 76: 1209-1212.
- Turkensteen L.J. 1978. Phytophthora infestans: three new hosts and specialized form causing
- foliar blight of *Solanum muricatum* in Peru. Plant Dis. Reptr. 62: 829. Van der Vossen E., Sikkema A., Hekkert B.t.L., Gros J., Stevens P., Muskens M., Wouters D., Pereira A., Stiekema W., Allefs S. 2003. An ancient R gene from the wild species *Solanum* bulbocastanum confers broad-spectrum resistance to Phytophthora infestans in cultivated potato and tomato. Plant J. 36: 867-882.

Wastie R.L. 1991. Breeding for resistance. (In:) Advanced Plant Pathology; *Phytophthora infestans*, the cause of late blight of potato. Ingram D.S., Williams P.H. (eds), Vol. 7. Academic Press, London, UK: 193–224.
Zwankhuizen M.J., Govers F., Zadoks J.C. 1998. Development of potato late blight epidemics: disease foci, disease gradients, and infection sources. Phytopathology 88: 754–763.